

Analysis of Germline Variants with Low Allele Frequencies in Turkish Healthy Cohort

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Abstract

BACKGROUND/AIMS: Within a specific population, rare variants with minor allele frequencies (MAFs) below 0.01% may underlie a particular genetic disorder when Mendelian diseases are investigated. Although MAFs can be determined by universal databases, the importance of a variant with a low MAF may vary across populations. Hence, documentation of rare variants in healthy individuals could be remarkably valuable. However, such approaches have not yet been applied in the Turkish population. Thus, we aimed to identify rare germline variants in a healthy Turkish population.

MATERIALS AND METHODS: We re-analyzed whole-exome sequencing data from 80 healthy Turkish individuals and filtered variants using a MAF threshold of <0.01%. We further assessed the pathogenicity of the filtered variants according to the American College of Medical Genetics criteria.

RESULTS: There were numerous rare variants, some of which were common to all participants. Importantly, those variants were classified as of unknown significance or as likely pathogenic; however, this classification should be revised because the variants were observed in healthy individuals.

CONCLUSION: We propose that these variants could be benign, as they were detected in healthy Turkish participants. Finally, the rare variants in the Turkish population should still be reported to guide clinicians during routine molecular diagnosis.

Keywords: Exome sequencing, Turkish population, rare variants, minor allele frequencies

INTRODUCTION

Molecular causes of genetic disorders have been identified using genetic variants. Although many variants have been reported for specific genetic disorders, additional variants remain to be discovered.¹ Thus, studies in human genetics focus on exploring variants associated with, or resulting in, a particular phenotype.² Here, using the single-gene single-phenotype approach, Mendelian diseases are generally diagnosed by detecting rare variants in a population.³

Rare variants are defined as deoxyribonucleic acid (DNA) alterations with minor allele frequencies (MAFs) of less than 0.01% in a specific population.⁴ According to the American College of Medical Genetics (ACMG) criteria⁵, a variant with low allelic frequency is classified with moderate piece of evidence for pathogenicity (PM2). Thus, low MAFs could be informative for evaluating the unreported variant. However, the MAF cutoff may vary between populations.⁶ Moreover, unreported or not molecularly proven variants, even those with low MAFs, are classified as variants of unknown significance (VUS), which limits molecular diagnosis.⁷

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To determine the pathogenicity of a variant, population databases are required. Although global databases such as gnomAD⁸, ExAc⁹, and 1000 Genomes Project¹⁰ are available, some countries have established population-specific databases to reduce biases arising from population-based differences.¹¹⁻¹³ Similarly, Türkiye has reported a whole-exome and whole-genome database¹⁴ and launched a portal for variant search (<https://tgd.tuseb.gov.tr/>) based on data from healthy participants. Nonetheless, the portal allows only single-variant searches, restricting cumulative evaluation of variants with low MAFs. Moreover, some variants of interest were not listed in the database. Hence, a more comprehensive report on variants with low frequencies in healthy cohorts is still needed.

In the present study, we re-analyzed whole exome sequencing (WES) data from 80 healthy Turkish individuals with respect to rare variants. After filtering the variants according to MAF values (<0.01%) from universal databases, we used the Franklin by Genoox tool (<https://franklin.genoox.com/>) to clarify variant pathogenicity according to ACMG criteria and listed the non-benign variants. Our results indicated that numerous variants had low MAFs, including variants in critical genes that were classified as VUS or likely pathogenic (LP). However, the proportion of those variants in the healthy cohort led to the conclusion that those variants, especially those linked to early-onset genetic disorders, should be re-categorized as benign in the Turkish population. The present study is the first to underscore the importance of rare variants in healthy individuals in the Turkish population, and such studies are still required.

MATERIALS AND METHODS

Ethics and Participants

In the present study, the WES data previously obtained from 80 healthy individuals in the sport genetics studies^{15,16} were re-analyzed. In previous studies, only polymorphisms of the participants were evaluated using an exome-wide association study. Sixty participants were elite Turkish athletes, and twenty were healthy, unrelated Turkish individuals. In the present study, all participants (27 females, 33.75%; 53 males, 66.25%; age >18 years) were regarded as healthy individuals and were not grouped, as none had any known (declared) genetic disorders. The study was approved by the Gazi University Non-Interventional Clinical Research Ethics Committee (approval number: 09, date: 05.04.2021). Under this ethical approval, all variants were already approved for analysis. Both written and verbal consents were obtained from the participants before the study. Moreover, the data were publicly available at <https://doi.org/10.6084/m9.figshare.24496216.v1>.

Variant Prioritization

In the study, data generated by WES, which was performed using the Twist Human Comprehensive Exome Panel (Twist Biosciences, USA) and the Illumina NextSeq500 (Illumina Inc., USA) on genomic DNAs from peripheral blood samples of the participants, were further examined. For the analysis of rare variants, the variants in variant call format for each participant were annotated using the ANNOVAR tool¹⁷ with the hg19 human reference genome. Next, variants located in exons or exon-intron boundaries were prioritized using VarAFT software¹⁸, where variants with read depth >10 were filtered by MAF <0.01%, considering only morbid genes.¹⁹ To limit the number of variants and to avoid manipulating any possible *de novo* variants, only homozygous variants were evaluated, while hemizygous or homozygous variants on chromosome X were documented regardless of participant sex.

Finally, the pathogenicity of the variants was determined using the Franklin by Genoox tool (<https://franklin.genoox.com/>) according to the ACMG criteria.⁵ Moreover, we queried the Turkish Genome Project (TGP) database (TGP; <https://tgd.tuseb.gov.tr/>) for each variant, using a single-variant search when the variant was listed, and excluded variants reported in the database.

Statistical Analysis

No statistical analysis was performed in the study.

RESULTS

Each participant had nearly 700,000 variants before filtering. Applying a MAF <0.001 filter to homozygous variants in morbid genes yielded approximately 15 variants per patient. Variant assessment using Franklin by Genoox according to the ACMG criteria showed approximately three variants per patient classified as VUS or LP; on average, one such variant per patient was not documented in the TGP database. Those variants listed in Table 1 were detected in at least one participant. Variants that met ACMG criteria BS2 (observed in a homozygous state in population databases more than expected for disease), BS3 (well-established functional studies show no damaging effect on protein function or splicing), BP3 (in-frame deletions/insertions in a repetitive region with no known function), or BP6 (reported as benign but lacking evidence for independent laboratory evaluation), as well as variants classified as VUS by the Franklin tool were not reported in the results.

DISCUSSION

In the present study, we re-analyzed the WES data from 80 healthy Turkish individuals and documented homozygous rare variants that were recurrently observed in the participants and not reported in the TGP database. Regarding the early-onset status of diseases linked to genes harboring detected variants, we documented 62 variants in 60 genes associated with diverse genetic disorders.

Rare variants are regarded as absent from control (healthy) individuals, and their frequencies are extremely low in population-based genome databases.²⁰ Thus, they are annotated with moderate evidence of pathogenicity (PM2) according to the ACMG criteria (5). Even though marking a variant with the PM2 criterion is suggestive of pathogenicity, studies have pointed out benign or likely benign variants in the presence of the PM2 criterion.²¹ Hence, reporting rare variants in healthy individuals may be useful to determine whether variants meeting the PM2 criterion are benign or likely benign. In the present cohort, 60 variants were annotated with the PM2 criterion, indicating that their frequencies were markedly low in public genome databases.

The frequency of a variant of interest can be determined using various genomic databases. Nonetheless, those databases are population-specific, and the exact frequencies vary across populations.²² Recently, Türkiye initiated a genome project called TGP, containing genomic information from 557 so-called healthy individuals. The platform allows users to search for individual variants of interest and provides a valuable opportunity to determine variant frequencies in a manner specific to the Turkish population. However, we noted that the platform does not document variants on the sex chromosomes, particularly indel variants. Therefore, listing rare variants in healthy Turkish individuals is fundamental to the assessment of variant pathogenicity in medical genetics in Türkiye.

Table 1. List of the rare variants with VUS or LP classification and not reported in the TGP database in 80 healthy participants

[illegible]

Table 1. Continued					
Gene	Variant	Depth/allele frequency	Pathogenicity	ACMG Criteria	Disease, inheritance pattern (OMIM#)
<i>GCM2</i>	NM_004752.4: c.283G>C: p.Asp95His	53/1.0	VUS	PM2	-Hyperparathyroidism 4, AD (617343) -Hypoparathyroidism, familial isolated 2, AD/AR (618883)
<i>GFI1</i>	NM_005263.5: c.925-3_925-2insTCTCTC	12/0.92	VUS	PM2	-?Neutropenia, nonimmune chronic idiopathic, of adults, AD (607847) -Neutropenia, severe congenital 2, AD (613107)
<i>HTT</i>	NM_002111.8:c.51_52insAGCAGCAGCAGCAGC: p.Phe17_Gln18insSerSerSerSerSer	74/1.0	VUS	PM2, PM4	-Huntington disease, AD (143100) -Lopes-Maciel-Rodan syndrome, AR (617435)
<i>INPP5E</i>	NM_019892.6:c.1159+8_1159+9insTGGCTGGAGGGGTGGGCG	28/1.0	VUS	PM2	-Impaired intellectual development, truncal obesity, retinal dystrophy, and micropenis syndrome, AR (610156) -Joubert syndrome 1, AR (213300)
<i>INTU</i>	NM_015693.4: c.1091+6A>C	32/1.0	VUS	PM2, BP4	-?Orofaciodigital syndrome XVII, AR (617926) -Short-rib thoracic dysplasia 20 with polydactyly, AR (617925)
<i>JPH3</i>	NM_001271604.4: c.430_431insTGCTGC: p.Leu143_Pro144insLeuLeu	104/0.98	VUS	PM2, PM4	-Huntington disease-like 2, AD (606438)
<i>KCNN3</i>	NM_002249.6: c.243_244insGCAGCAGCAGCAGCA: p.Pro81_Pro82insAlaAlaAlaAlaAla	93/1.0	VUS	PM2, PM4	-Zimmermann-Laband syndrome 3, AD (618658)
<i>KDM5C</i>	NM_004187.5:c.2517-6_2517-5insACT	22/1.0	VUS	PM2	-Intellectual developmental disorder, X-linked syndromic, Claes-Jensen type, XLR (300534)
<i>KDM6B</i>	NM_001348716.2:c.751_752insACCACC: p.Pro250_Leu251insTyrHis	59/0.95	VUS	PM2, BP3	-Stolerman neurodevelopmental syndrome, AD (618505)
<i>KIF4A</i>	NM_012310.5: c.1924A>G: p.Met642Val	22/1.0	VUS	PM2, PM4	-Intellectual developmental disorder, X-linked 100, XLR (300923) -Taurodontism, microdontia, and dens invaginatus, XLR (313490)
<i>KLHL15</i>	NM_030624.3: c.1805G>A: p.Arg602His	22/1.0	VUS	PM2, PP2	-Intellectual developmental disorder, X-linked 103, XLR (300982)
<i>KRT10</i>	NM_000421.5: c.1684_1685insAGCTCCGGCGGCGGATACGGCGGCGGCAGC: p.Ser562*	34/1.0	LP	PVS1, PM2	-?Ichthyosis histrix, Lambert type, AD (146600) -Epidermolytic hyperkeratosis 2A, AD (620150) -Epidermolytic hyperkeratosis 2B, AR (620707) -Ichthyosis with confetti, AD (609165) -Ichthyosis, annular epidermolytic 1, AD (607602)
<i>LFNG</i>	NM_001166355.2:c.137_138insGATG: p.Asp47Metfs*11	83/1.0	VUS	PM2	-Spondylocostal dysostosis 3, AR (609813)
<i>LRSAM1</i>	NM_001005373.4: c.593C>T: p.Ala198Val	67/1.0	VUS	PM2, BP4	-Charcot-Marie-Tooth disease, axonal, type 2P, AD/AR (614436)
<i>MLC1</i>	NM_015166.4: c.1053_1054insCGGGGAGGTGAGTGGCTGTGGGGTGGGGGTGC: p.Ala351_Gly352insArgGlyGlyGluTrpProValGlyTrpGlyCys	41/1.0	VUS	PM2, PM4	-Megalencephalic leukoencephalopathy with subcortical cysts 1, AR (604004)
<i>NEFH</i>	NM_021076.4:c.1939_1940insGTCCCTGAGAAGGCCAA:p.Ala646_Lys647insSerProLeuArgArgPro	95/0.97	VUS	PM2, BP4	-?Amyotrophic lateral sclerosis, susceptibility to, AD, AR (105400) -Charcot-Marie-Tooth disease, axonal, type 2CC, AD (616924)
<i>OGT</i>	NM_181672.3: c.922A>G: p.Ser308Gly	33/0.97	VUS	PM2, PP2	-Intellectual developmental disorder, X-linked 106, XLR (300997)
<i>OTC</i>	NM_000531.6: c.76C>T: p.Arg26Trp	29/1.0	VUS	PM2, PM5, PP2	-Ornithine transcarbamylase deficiency, XL (311250)

Table 1. Continued					
Gene	Variant	Depth/allele frequency	Pathogenicity	ACMG Criteria	Disease, inheritance pattern (OMIM#)
<i>OTOG</i>	NM_001292063.2: c.89G>A: p.Arg30His	57/1.0	VUS	PM2, BP4	-Deafness, autosomal recessive 18B, AR (614945)
<i>PAPPA2</i>	NM_020318.3: c.2162A>G: p.Tyr721Cys	32/1.0	VUS	PM2	-Short stature, Dauber-Argente type, AR (619489)
<i>PAPSS2</i>	NM_001015880.2: c.381+1_381+2insAAAAA	12/1.0	VUS	PVS1	Brachyolmia 4 with mild epiphyseal and metaphyseal changes, AR (612847)
<i>PCNT</i>	NM_006031.6: c.427_428insTGGGATGTTACAGTCAGTGACCCACCAGAACAA: p.Arg143delinsLeuGlyCysSerGlnSerValThrThrHisGlnAsnSerSer	24/1.0	VUS	PM2, PM4	-Microcephalic osteodysplastic primordial dwarfism, type II, AR (210720)
<i>POGZ</i>	NM_015100.4: c.3606C>G: p.Asp1202Glu	64/1.0	VUS	PM2, PP2	-White-Sutton syndrome, AD (616364)
<i>POLR3A</i>	NM_007055.4: c.3503A>C: p.His1168Pro	60/1.0	VUS	PM2, PP2	-Leukodystrophy, hypomyelinating, 7, with or without oligodontia and/or hypogonadotropic hypogonadism, AR (607694) -Wiedemann-Rautenstrauch syndrome, AR (264090)
<i>POU3F4</i>	NM_000307.5: c.517G>C: p.Gly173Arg	34/1.0	VUS	PM2	-Deafness, X-linked 2, XLR (304400)
<i>PPFIBP1</i>	NM_003622.4: c.2764A>G: p.Met922Val	72/1.0	VUS	PM2	-Neurodevelopmental disorder with seizures, microcephaly, and brain abnormalities, AR (620024)
<i>RBM10</i>	NM_005676.5: c.1166T>C: p.Met389Thr	34/0.97	VUS	PM2, PP2, BP4	-TARP syndrome, XLR (311900)
<i>SEC63</i>	NM_007214.5: c.1936-6_1936-5dup	18/94	VUS	-	-Polycystic liver disease 2, AD (617004)
<i>SH3BP2</i>	NM_001122681.2: c.1507G>A: p.Glu503Lys	52/1.0	VUS	PM2	-Cherubism, AD (118400)
<i>SLC12A2</i>	NM_001046.3: c.284_285insGCGGCGGCGGCG: p.Ala95_Ala96insArgArgArgArg	14/0.93	VUS	PM2, PM4	-Deafness, autosomal dominant 78, AD (619081) -Delpire-McNeill syndrome, AD (619083) -Kilquist syndrome, AR (619080)
<i>SLC4A10</i>	NM_001178016.2: c.81+2T>C	49/1.0	VUS	PM2	-Neurodevelopmental disorder with hypotonia and characteristic brain abnormalities, AR (620746)
<i>SLC44A4</i>	NM_025257.3: c.845T>C: p.Val282Ala	73/1.0	VUS	PM2, BP4	-?Deafness, autosomal dominant 72, AD (617606)
<i>SLC9A6</i>	NM_001379110.1: c.1581+8T>A	32/1.0	VUS	PM2	-Intellectual developmental disorder, X-linked syndromic, Christianson type, XL (300243)
<i>SMC1A</i>	NM_006306.4: c.1911+3G>A	54/1.0	VUS	PM2, BP4	-Cornelia de Lange syndrome 2, XLD (300590) -Developmental and epileptic encephalopathy 85, with or without midline brain defects, XLD (301044)
<i>SMS</i>	NM_004595.5: c.536G>A: p.Arg179Gln	66/1.0	VUS	PM2, PP2	-Intellectual developmental disorder, X-linked syndromic, Snyder-Robinson type, XLR (309583)
<i>SPNS2</i>	NM_001124758.3: c.1607+5dup	43/1.0	VUS	PM2, PP3	-?Deafness, autosomal recessive 115, AR (618457)
<i>SV2A</i>	NM_014849.5: c.730G>T: p.Val244Phe	57/1.0	VUS	PM2	-Developmental and epileptic encephalopathy 113, AR (620772)
<i>TTC8</i>	NM_144596.4: c.909G>C: p.Glu303Asp	51/1.0	VUS	PM2, PP3	-?Retinitis pigmentosa 51, AR (613464) -Bardet-Biedl syndrome 8, AR (615985)

Table 1. Continued					
Gene	Variant	Depth/allele frequency	Pathogenicity	ACMG Criteria	Disease, inheritance pattern (OMIM#)
USP9X	NM_001039591.3: c.3496C>T: p.His1166Tyr	21/1.0	VUS	PM2, PP2	-Intellectual developmental disorder, X-linked 99, XLR (300919) -Intellectual developmental disorder, X-linked 99, syndromic, female-restricted, XLD (300968)
	NM_020922.5: c.4870+8T>A	25/1.0	VUS	PM2	-Prieto syndrome, XLR (309610)
WNK3	NM_020922.5: c.3652-8_3652-7insTTGTT	13/1.0	VUS	PM2	
ZNF711	NM_001330574.2: c.547A>G: p.Thr183Ala	36/1.0	VUS	PM2, BP4	-Intellectual developmental disorder, X-linked 97, XL (300803)
VUS: Variants of unknown significance, LP: Likely pathogenic, AD: Autosomal dominant, AR: Autosomal recessive, XLR: X-linked recessive, XLD: X-linked dominant, ACMG: American College of Medical Genetics, OMIM: Online Mendelian Inheritance in Man, PVS1: Pathogenic very strong 1.					

Among the rare variants detected in the healthy cohort, two variants (*FAM20C*: NM_020223.4:c.951_952insACAGGTGAGCCCTTCCTCCTCCATCCGCG:p.Asp318Thrfs*118 and *KRT10*:NM_000421.5:c.1684_1685insAGCTCCGGCGGCGGATACGGCGGCGGCAGC:p.Ser562*) were classified as LP based on PM2 and the very strong pathogenic very strong 1 (PVS1) criteria for pathogenicity. The PVS1 criterion applies to variant types that result in loss of protein function, including frameshifts, altered splicing, and exonic deletions.²³ Those variants were detected in at least three healthy participants in the Turkish population. *FAM20C* (Golgi-associated secretory pathway kinase) encodes a protein that functions as a Golgi casein kinase, regulates phosphorylation of secreted proteins, and is linked to Raine Syndrome (Online Mendelian Inheritance in Man #259775), which is characterized by neonatal osteosclerotic bone dysplasia.²⁴ However, the LP variant in the *FAM20C* gene was detected in five independent healthy individuals in the Turkish population, and therefore is unlikely to be the cause of lethal Raine Syndrome. Interestingly, the frameshift variant localizes to the kinase domain of the protein.²⁵ Thus, further molecular studies are required to determine how protein function is conserved in the presence of this variant. The other variant in *KRT10* was observed in three participants. The *KRT10* gene encodes a type I keratin that is mainly expressed in epidermal cells. The mutations in this gene have been associated with rare skin anomalies. Importantly, the C-terminus of the protein was identified as a mutation hotspot.^{26,27} According to the ENSEMBL database²⁸, *KRT10* gene (NM_000421.5) encodes a 584 amino acid-length protein. Hence, the variant detected in the healthy participants was localized to the end of the protein, which may not affect the protein’s structure and function.²⁹

During classification, unreported or unproven variants are classified as VUS.³⁰ In the present study, 60 variants were classified as VUS. The classification would change when the variant is detected in healthy individuals.³¹ Therefore, we propose that the classification of those variants be re-evaluated in medical genetics in Türkiye.

In the present study, we report biallelic variants identified in healthy individuals. Among the 60 genes affected in the homozygous state, 14 have been linked exclusively to dominant genetic disorders. In specific cases, the homozygous variants in dominant diseases have been reported to be neutral³², further proving the non-pathogenic nature of the listed variants.

Study Limitations

Although the study submitted variant lists for a limited number of healthy individuals, similar studies with larger group of participants or a wider sequencing approach as whole genome sequencing should be conducted to define more variants that would be reclassified in Turkish population.

CONCLUSION

The present study documented rare germline and homozygous variants in genes linked to early-onset genetic disorders in healthy Turkish individuals. Although the variants were classified as VUS or LP, their detection in the healthy cohort may indicate that those variants are benign. Moreover, the listed variants were not reported in the TGP database. Thus, reporting rare variants in a population-specific manner, as in the present study, is still valuable for guiding medical geneticists when prioritizing variants. Nevertheless, possible multigenic or multifactorial inheritance patterns should be considered when referring to those variants.

MAIN POINTS

- Rare variants are common in Turkish healthy cohort.
- The reported rare variants of unknown significance may be reclassified as benign in the Turkish population.
- Further population-based studies could facilitate variant prioritization.

ETHICS

Ethics Committee Approval: The study was approved by the Gazi University Non-Interventional Clinical Research Ethics Committee (approval number: 09, date: 05.04.2021).

Informed Consent: Informed consent was obtained from all subjects involved in the study.

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Footnotes

Authorship Contributions

Concept: S.G.E., C.B., H.H.K., Design: S.G.E., G.A., C.B., H.H.K., Data Collection and/or Processing: C.B., H.H.K., Analysis and/or Interpretation: S.G.E., F.Y.T., G.A., H.H.K., Writing: S.G.E., F.Y.T., G.A., C.B., H.H.K.

DISCLOSURES

Conflict of Interest: No conflict of interest was declared by the authors.

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REFERENCES

- Saint Pierre A, Génin E. How important are rare variants in common disease? *Brief Funct Genomics*. 2014; 13(5): 353-61.
- Johnston HR, Hu Y, Cutler DJ. Population genetics identifies challenges in analyzing rare variants. *Genet Epidemiol*. 2015; 39(3): 145-8.
- Auer PL, Lettrec G. Rare variant association studies: considerations, challenges and opportunities. *Genome Med*. 2015; 7(1): 16.
- Goswami C, Chattopadhyay A, Chuang EY. Rare variants: data types and analysis strategies. *Ann Transl Med*. 2021; 9(12): 961.
- Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. *Genet Med*. 2015; 17(5): 405-24.
- Shearer AE, Eppsteiner RW, Booth KT, Ephraim SS, Gurrola J 2nd, Simpson A, et al. Utilizing ethnic-specific differences in minor allele frequency to recategorize reported pathogenic deafness variants. *Am J Hum Genet*. 2014; 95(4): 445-53.
- Wang J, Shen Y. When a "disease-causing mutation" is not a pathogenic variant. *Clin Chem*. 2014; 60(5): 711-3.
- Gudmundsson S, Singer-Berk M, Watts NA, Phu W, Goodrich JK, Solomonson M, et al. Variant interpretation using population databases: lessons from gnomAD. *Hum Mutat*. 2022; 43(8): 1012-30.
- Kobayashi Y, Yang S, Nykamp K, Garcia J, Lincoln SE, Topper SE. Pathogenic variant burden in the ExAC database: an empirical approach to evaluating population data for clinical variant interpretation. *Genome Med*. 2017; 9(1): 13.
- Clarke L, Zheng-Bradley X, Smith R, Kulesha E, Xiao C, Toneva I, et al. The 1000 Genomes Project: data management and community access. *Nat Methods*. 2012; 9(5): 459-62.
- Johnston JJ, Biesecker LG. Databases of genomic variation and phenotypes: existing resources and future needs. *Hum Mol Genet*. 2013; 22(R1): R27-31.
- Higasa K, Miyake N, Yoshimura J, Okamura K, Niihori T, Saitsu H, et al. Human genetic variation database, a reference database of genetic variations in the Japanese population. *J Hum Genet*. 2016; 61(6): 547-53.
- Lee S, Seo J, Park J, Nam JY, Choi A, Ignatius JS, et al. Korean Variant Archive (KOVA): a reference database of genetic variations in the Korean population. *Sci Rep*. 2017; 7(1): 4287.
- Correction for Kars et al., The genetic structure of the Turkish population reveals high levels of variation and admixture. *Proc Natl Acad Sci U S A*. 2021; 118(52): 2120031118. Erratum for: *Proc Natl Acad Sci U S A*. 2021; 118(36): e2026076118.
- Bulgay C, Kasakolu A, Kazan HH, Mijaica R, Zorba E, Akman O, et al. Exome-wide association study of competitive performance in elite athletes. *Genes (Basel)*. 2023; 14(3): 660.
- Kazan HH, Kasakolu A, Koncagul S, Ergun MA, John G, Sultanov RI, et al. Association analysis of indel variants and gene expression identifies MDM4 as a novel locus for skeletal muscle hypertrophy and power athlete status. *Exp Physiol*. 2025; 110(11): 1661-71.
- Wang K, Li M, Hakonarson H. ANNOVAR: functional annotation of genetic variants from high-throughput sequencing data. *Nucleic Acids Res*. 2010; 38(16): e164.
- Desvignes JP, Bartoli M, Delague V, Krahn M, Miltgen M, Bérout C, et al. VarAFT: a variant annotation and filtration system for human next generation sequencing data. *Nucleic Acids Res*. 2018; 46(W1): W545-53.
- Jauss RT, Popp B, Bachmann J, Abou Jamra R, Platzer K. The MorbidGenes panel: a monthly updated list of diagnostically relevant rare disease genes derived from diverse sources. *Hum Genet*. 2024; 143(12): 1459-63.
- Davieson CD, Joyce KE, Sharma L, Shovlin CL. DNA variant classification-reconsidering "allele rarity" and "phenotype" criteria in ACMG/AMP guidelines. *Eur J Med Genet*. 2021; 64(10): 104312.
- Qu HQ, Wang X, Tian L, Hakonarson H. Application of ACMG criteria to classify variants in the human gene mutation database. *J Hum Genet*. 2019; 64(11): 1091-5.
- International HapMap 3 Consortium; Altshuler DM, Gibbs RA, Peltonen L, Altshuler DM, Gibbs RA, et al. Integrating common and rare genetic variation in diverse human populations. *Nature*. 2010; 467(7311): 52-8.
- Abou Tayoun AN, Pesaran T, DiStefano MT, Oza A, Rehm HL, Biesecker LG, et al. Recommendations for interpreting the loss of function PVS1 ACMG/AMP variant criterion. *Hum Mutat*. 2018; 39(11): 1517-24.
- Koob M, Doray B, Fradin M, Astruc D, Dietemann JL. Raine syndrome: expanding the radiological spectrum. *Pediatr Radiol*. 2011; 41(3): 389-93.
- Xu R, Tan H, Zhang J, Yuan Z, Xie Q, Zhang L. Fam20C in human diseases: emerging biological functions and therapeutic implications. *Front Mol Biosci*. 2021; 8: 790172.
- Tsubota A, Akiyama M, Kanitakis J, Sakai K, Nomura T, Claudy A, et al. Mild recessive bullous congenital ichthyosiform erythroderma due to a previously unidentified homozygous keratin 10 nonsense mutation. *J Invest Dermatol*. 2008; 128(7): 1648-52.
- Takeichi T, Suga Y, Mizuno T, Okuno Y, Ichikawa D, Kono M, et al. Recurrent KRT10 variant in ichthyosis with confetti. *Acta Derm Venereol*. 2020; 100(14): adv00209.
- Dyer SC, Austine-Orimoloye O, Azov AG, Barba M, Barnes I, Barrera-Enriquez VP, et al. Ensembl 2025. *Nucleic Acids Res*. 2025; 53(D1): D948-57.
- Hu J, Ng PC. Predicting the effects of frameshifting indels. *Genome Biol*. 2012; 13(2): R9.
- Fowler DM, Rehm HL. Will variants of uncertain significance still exist in 2030? *Am J Hum Genet*. 2024; 111(1): 5-10.
- Weck KE. Interpretation of genomic sequencing: variants should be considered uncertain until proven guilty. *Genet Med*. 2018; 20(3): 291-3.
- Monies D, Maddirevula S, Kurdi W, Alanazy MH, Alkhalidi H, Al-Owain M, et al. Autozygosity reveals recessive mutations and novel mechanisms in dominant genes: implications in variant interpretation. *Genet Med*. 2017; 19(10): 1144-50.